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(64) **19-Nor-vitamin D3 compounds with substituent at 2-position.**

(57) The 2α and 2β -hydroxy as well as the $2\alpha(3'$ -hydroxypropoxy)- and $2\beta(3'$ -hydroxypropoxy)- and $2\alpha(\text{benzyloxy})$ - analogs of 19-nor- $1\alpha,25$ -dihydroxyvitamin D_3 are disclosed. The two 2-hydroxy analogs showed in vivo calcium transport with little or no bone calcium mobilization; the 2β -more than the 2α -analog. Both analogs induced differentiation of malignant cells. The two analogs thus show promise in the treatment of osteoporosis. The 2α -hydroxypropoxy analog showed a selective activity profile, combining high potency in inducing differentiation of malignant cells with very low or no bone calcification activity, a possible use in the treatment of malignancies.

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This invention relates to biologically active vitamin D₃ compounds. More specifically, the invention relates to 19-nor-analogs of 1 α -hydroxylated vitamin D₃ compounds having a substituent at the 2-position in the A-ring.

5 **Background and Summary of the Invention**

The 1 α -hydroxylated metabolites of vitamin D -- most importantly 1 α ,25-dihydroxyvitamin D₃ and 1 α ,25-dihydroxyvitamin D₂ -- are known as highly potent regulators of calcium homeostasis in animals and humans, and more recently their activity in cellular differentiation has also been established. V. Ostrem et al, Proc. Natl. Acad. Sci. USA, (1987), 84, 2810. As a consequence, many structural analogs of these metabolites, such as compounds with different side chain structures, different hydroxylation patterns, or different stereochemistry, have been prepared and tested. Important examples of such analogs are 1 α -hydroxyvitamin D₃, 1 α -hydroxyvitamin D₂, various side chain fluorinated derivatives of 1 α ,25-dihydroxyvitamin D₃, and side chain homologated analogs. Several of these known compounds exhibit highly potent activity *in vivo* or *in vitro*, and some of these have been found to exhibit an interesting separation of activities in cell differentiation and calcium regulation. This difference in activity provides these compounds with advantageous therapeutic activity profiles and thus numerous of these compounds are in use, or have been proposed for use, in the treatment of a variety of diseases such as renal osteodystrophy, vitamin D-resistant rickets, osteoporosis, psoriasis, and certain malignancies.

Recently, a new class of vitamin D analogs has been discovered, i.e. the so-called 19-nor-vitamin D compounds. 19-Nor-vitamin D compounds are vitamin D analogs in which the ring A exocyclic methylene group (carbon 19) typical of all vitamin D compounds has been removed and replaced by two hydrogen atoms. Specifically, these compounds exhibit a selective activity profile with high potency in inducing cellular differentiation, and minimal bone calcification activity. Such a differential activity profile renders these compounds useful for the treatment of malignancies, or the treatment of various skin disorders. Two different methods of synthesis of these 19-nor-vitamin D analogs have been described (Perlman et al. Tetrahedron Letters 31, 1823 (1990); Perlman et al Tetrahedron Letters 32, 7883 (1991), and DeLuca et al U.S. Patent 5,086,191).

In U.S. Patent 4,666,634, 2 β 3-hydroxy and alkoxy analogs of 1 α ,25-dihydroxyvitamin D₃ have been described and examined as potential drugs for osteoporosis and as antitumor agents. See also T. Okano et al, Biochem. and Biophys. Res. Comm., (1989) 163, 1444. However the new analogs also have many undesired side effects most notable of which is the potential development of hypercalcemia upon administration.

In a continuing effort to explore the new 19-nor class of pharmacologically important vitamin D analogs, the 2 α - and 2 β -hydroxy as well as the 2 α (3'-hydroxypropoxy)- and the 2 β (3'-hydroxypropoxy)- and 2 α (benzyloxy)-analogs of 19-nor-1 α ,25-dihydroxyvitamin D₃ have now been synthesized. The two 2-hydroxy analogs showed *in vivo* calcium transport with little or no bone calcium mobilization; the 2 β - more than the 2 α - analog. Both 2-hydroxy analogs induced differentiation of malignant cells. These two analogs thus show promise in the treatment of osteoporosis. The 2 α -hydroxypropoxy analog showed a selective activity profile, combining high potency in inducing differentiation of malignant cells with very low bone calcium mobilizing activity, a possible use in the treatment of cancer.

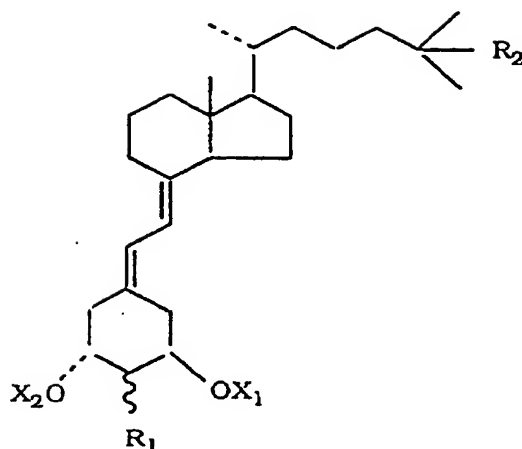
40 **Brief Description of the Drawings**

Figs. 1a and 1b are graphs of the percent differentiation of HL-60 cells versus concentration for a prior art vitamin D₃ compound and three of the new 19-nor-vitamin D₃ compounds; and

Figs. 2a and 2b are graphs of the competitive binding ability versus concentration for the same four compounds as in Figs. 1a and 1b.

50 **Disclosure of the Invention**

The new 19-nor-Vitamin D₃ analogs having a substituent at the 2-position in the A-ring are represented by the following general formula:



where X_1 and X_2 are each selected from hydrogen or a hydroxy protecting group, R_1 which may be in either α or β position is a hydroxy group, protected hydroxy group or the group OR_3 where R_3 is an alkyl, hydroxyalkyl, fluoroalkyl, arylalkyl or aryl group, and R_2 is hydrogen or a hydroxy group. Specific examples of the compounds represented by the formula above are: 1 α ,2 α ,25-trihydroxy-19-nor-vitamin D_3 ; 1 α , 2 β , 25-trihydroxy-19-nor-vitamin D_3 ; 1 α ,25-dihydroxy-2 α -(3'-hydroxypropoxy)-19-nor-vitamin D_3 ; 1 α ,25-dihydroxy-2 β -(3'-hydroxypropoxy)-19-nor-vitamin D_3 and 1 α ,25-dihydroxy-2 α -(benzyloxy)-19-nor-vitamin D_3 .

The starting material for the synthesis of these new 19-nor compounds is the commercially available synthon (1R,3R,4R,5R) (-) quinic acid, D. Desmaele et al, Tetrahedron Letters (1985) 26, 4941, which after esterification with methanol in the presence of catalytic amount of p-toluene sulfonic acid, followed by treatment with tert-butyltrimethylsilyl chloride and triethylamine in dimethyl formamide gave the protected methyl ester 1. Reduction with diisobutylaluminum hydride gave polyalcohol 2 followed by sodium periodate oxidation to the cyclohexanone derivative 3. The 4-hydroxy group was protected with N-(trimethylsilyl) imidazole to give 4.

The trimethylsilyl protecting group in this particular case with the very hindered 2-hydroxy group gave the needed chemoselectivity in the 2 position throughout all subsequent steps. Peterson reaction with methyl (trimethylsilyl)acetate in the presence of LDA in anhydrous tetrahydrofuran gave a mixture of the protected cyclohexylidene esters 5a, 5b. The latter was reduced to the allylic alcohols 6a, 6b with diisobutylaluminum hydride and separated and finally transformed to the desired phosphine oxide 7a and 7b by in situ conversion to the tosylate with BuLi and tosylchloride followed by lithium diphenylphosphide and oxidation with hydrogen peroxide. (During all these steps the 2 trimethylsilyloxy group was not removed).

The synthesis of the CD ring Windaus Grundmann ketone with the appropriate protected side chain is well documented in the literature. See E. G. Baggiolini et al, J. Org. Chem. (1986) 51, 3098; F. J. Sardina et al, J. Org. Chem. (1986) 51, 1264; and U. S. Patent 4,804,502. In the present disclosure, the recently described procedure i.e. ozonolysis followed by RuO_4 oxidation of commercial vitamin D_3 was chosen. See J. Klegel et al, Tetrahedron Letters 32, 6057 (1991). The 25-OH of the Grundmann ketone was protected with TES-Cl, imidazole, in DMF to give chemoselectivity from the 2-OTMS group in the 19-nor product. With the required synthons on hand, the final convergent formation of 19-nor-1 α ,2 α and -2 β ,25-dihydroxy-cholecalciferol 9a, 10a was then accomplished. See B. Lythgoe et al, J. Chem. Soc. Perkin Trans. 1, (1978) 590; B. Lythgoe et al, J. Chem. Soc. Perkin Trans. 1, (1976) 2386; and H. T. Toh et al, J. Org. Chem. (1983), 48, 1414. Wittig Horner reaction of the lithium phosphinoxy carbanion prepared from 7a and n-butyl lithium at -78°C in tetrahydrofuran with the protected Windaus Grundmann ketone 8 proceeded to give the desired 19-nor-vitamin derivative 9a which after deprotection gave the crystalline 19-nor-1 α ,2 α ,25-trihydroxyvitamin D_3 10a; 10b was obtained in the same manner from 7b.

For the synthesis of the 19-nor-2 α -(3'-hydroxypropoxy)-1 α ,25-dihydroxyvitamin D_3 14a, 9a was partially hydrolyzed under carefully controlled conditions: i.e. 9a was treated with a mixture of 8:8:1 tetrahydrofuran acetic acid water at room temperature for 4.5 h and the resulting mixture separated by HPLC to give the free 2 α -hydroxy 11a. 3-bromo-1-tert-butyltrimethylsilyloxy propane 12 was chosen as the alkylating agent and was prepared by silylation from the corresponding bromo alcohol. 11a was treated with sodium hydride and the protected bromo compound 12 in the presence of 18-Crown-6 in anhyd. DMF for 48 h to give the protected 13a which

was deprotected with Bu_4NF in THF to give the expected 14a; 14b was obtained in the same manner from 11b. Under the same conditions 11a with benzylbromide gave after deprotection 16a.

The stereochemistry at the 2-position of 5 was determined first by separation of the enantiomers followed by partial hydrolysis, and acylation. The two enantiomer's structure was then determined by NOE followed by 2D NMR in deuterated benzene. This revealed that under the reaction conditions the major component was the 2 α -compound 5a and the minor the 2 β -compound 5b (3:1).

As used in the description, and in the claims, the term "hydroxy-protecting group" refers to any group commonly used for the protection of hydroxy functions during subsequent reactions, including, for example, acyl or alkylsilyl groups such as trimethylsilyl, triethylsilyl, t-butyltrimethylsilyl and analogous alkyl or arylsilyl radicals, or alkoxyalkyl groups such as methoxymethyl, ethoxymethyl, methoxyethoxymethyl, tetrahydrofuranyl or tetrahydropyranyl. A "protected-hydroxy" is a hydroxy function derivatized by one of the above hydroxy-protecting groupings. "Alkyl" represents a straight-chain or branched hydrocarbon radical of 1 to 10 carbons in all its isomeric forms, such as methyl, ethyl, propyl, isopropyl, butyl, isobutyl, pentyl, etc., and the terms "hydroxyalkyl," "fluoroalkyl" and "arylalkyl" refer to such an alkyl radical substituted by one or more hydroxy, fluoro or aryl groups respectively. An "acyl" group is an alkanoyl group of 1 to 6 carbons in all its isomeric forms, or an aroyl group, such as benzoyl, or halo-, nitro- or alkyl-substituted benzoyl groups, or an alkoxy-carbonyl group of the type alkyl-O-CO-, such as methoxycarbonyl, ethoxycarbonyl, propyloxycarbonyl, etc., or a dicarboxylic acyl group such as oxalyl, malonyl, succinoyl, glutaroyl, or adipoyl. The term "aryl" signifies a phenyl-, or an alkyl-, nitro- or halo-substituted phenyl group. The term alkoxy signifies the group alkyl-O-. This invention is more specifically described by the following illustrative examples. In these examples specific products identified by Arabic numerals (e.g. 1, 2, 3, etc.) refer to the specific structures so identified in the preceding description and in the Schemes.

Example 1

Preparation of 1 α ,2 α ,25-trihydroxy-19-nor-vitamin D₃ (10a) and 1 α ,2 β ,25-trihydroxy-19-nor-vitamin D₃ (10b).

(a) (1R,3R,4R,5R)(-) Methyl Quinicate.

Referring first to Scheme I, p-toluene sulfonic acid (0.5g) was added to a solution of quinic acid (12.74 g, 66.3 mmol) in methanol. The solution was stirred for 24 h. Solid NaHCO_3 (1.0g) was added and after 15 min the solution was filtered and concentrated to give 12.61 g (61.16 mmol) of the methyl ester in 92% yield.

(b) (1R,3R,4R,5R) Methyl 3,5-Bis(tert-butylidimethylsilyloxy), 1,4-dihydroxycyclohexane-carboxylate (1).

tert-Butylidimethylsilyl chloride (6.73 g, 44.62 mmol) was added to a solution of methyl (1R,3R,4R,5R)(-) quinicate (3.68 g, 17.85 mmol) and triethyl amine (6.2 mL, 44.62 mmol) in 44 mL of anhydrous dimethyl formamide at 0°C with stirring. After 4 h the solution was warmed to room temperature and stirring continued for another 14 h. The solution was poured into water and extracted with ether. The combined organic layers were extracted with brine, dried over anhydrous MgSO_4 , filtered and concentrated. The residue was purified by column chromatography on silica gel, eluting with 5-10% ethyl acetate in hexane mixtures, to give 4.6 g (60%) of 1 as a white solid. M.p. 82-82.5°C (after recrystallization from hexanes). ^1H NMR (CDCl_3 , 500 MHz) δ 4.53 (bs, 1 H), 4.36 (bs, 1 H), 4.11 (ddd, 1 H), 3.76 (s, 3 H), 3.42 (dd, 1 H), 2.31 (bs, 1 H), 2.18 (bd, 1 H), 2.05 (ddd, 2 H), 1.82 (dd, 1 H), 0.91 (s, 9 H), 0.89 (s, 9 H), 0.15 (s, 3 H), 0.14 (s, 3 H), 0.11 (s, 3 H), 0.09 (s, 3 H) MS m/e (relative intensity) 377 (70), 227 (91).

(c) (1R,3R,4R,5R) [3,5-Bis(tert-butylidimethylsilyloxy)-1,4-dihydroxy]-1-hydroxymethylcyclohexane (2).

Diisobutyl aluminum hydride (45mL, 45.0 mmol, 1.0 M in hexanes) was added to a solution of the ester (3.28 g, 7.5 mmol) in ether (45 mL) at -78°C. After 20 min. the solution was warmed to -23°C and stirred for 2h. The solution was diluted with ether and then 2N potassium sodium tartrate was slowly added. The solution was warmed to room temperature and stirred for 15 min. The ether layer was separated and the aqueous layer extracted with ether. The combined ether layers were extracted with brine, dried over anhydrous MgSO_4 , filtered and concentrated. The material was further purified by column chromatography on silica gel with 25% ethyl acetate/hexanes to give 83% of 2 (2.52 g, 6.20 mmol). Mp. 108-109°C from hexanes. ^1H NMR (CDCl_3 , 500 MHz) δ 4.52 (bs, 1 H), 4.32 (bs, 1 H), 4.12 (ddd, 1 H), 3.40 (dd, 1 H), 3.33 (dd, 2 H), 2.28 (d, 1 H), 2.11 (dd, 1 H), 2.00 (ddd, 2 H), 1.52 (dd, 1 H), 1.33 (dd, 1 H), 0.91 (s, 9 H), 0.90 (s, 9 H), 0.16 (s, 3 H), 0.14 (s, 3 H), 0.12 (s,

3 H), 0.11 (s, 3 H), MS m/e (relative intensity) : 349 (8), 331 (13), 239 (12), 199 (100).

(d) (3R,5R) [3,5-Bis(tert.-butyldimethylsilyloxy) 4-hydroxy]-1-cyclohexanone (3).

5 Sodium periodate saturated water (28.5 mL) was added to the triol **2** (1.91 g, 4.7 mmol) in methanol (124 mL) at 0°C. The solution was stirred for 1 h, then poured into water and extracted with ether. The combined ether fractions were washed with brine, dried over anhydrous MgSO₄, filtered and concentrated, to give 1.72 g (4.59 mmol) of **3** (98%). No further purification was required. Mp. 98-100°C from hexanes. ¹H NMR (CDCl₃, 500 MHz) δ 4.28 (m, 2 H), 3.80 (bs, 1 H), 2.77 (dd, 1 H, J=14.3, 3.4 Hz), 2.59 (dd, 1 H, J=13.1, 10.7 Hz), 2.45 (dd, 1 H, J=14.1, 5.2 Hz), 2.25 (bd, 1 H, J=15.9 Hz), 0.90 (s, 9 H), 0.85 (s, 9 H), 0.08 (s, 3 H), 0.08 (s, 3 H), 0.06 (s, 6 H), MS m/e (relative intensity) 317 (62), 231 (16), 185 (76), 143 (100).

(e) (3R,5R) [3,5-Bis(tert.-butyldimethylsilyloxy)-4-trimethylsilyloxy]-1-cyclohexanone (4).

15 N-(Trimethylsilyl)imidazole (2.52 mL, 26.67 mmol) was added to a solution of the ketoalcohol **3** (1.56 g, 4.167 mmol) in methylene chloride (38 mL). The solution was stirred for 20 h. Water (1 mL) was added and the solution stirred for 30 min. Brine and methylene chloride was added. The brine was extracted with methylene chloride. The combined methylene chloride fractions were dried with anhydrous MgSO₄, filtered and concentrated. The residue was further purified by column chromatography on silica gel with 10% ethyl acetate in hexane to give **4** (1.76 g, 3.95 mmol) in 95% yield. ¹H NMR (CDCl₃, 500 MHz) δ 4.25 (m, 1 H), 4.13 (m, 1 H), 4.04 (m, 1 H), 2.74 (ddd, 2 H), 2.38 (dd, 1 H), 2.19 (dd, 1 H), 0.90 (s, 9 H), 0.86 (s, 9 H), 0.16 (s, 9 H), 0.07 (bs, 12 H). MS m/e (relative intensity): 431 (5), 389 (100), 299 (45), 257 (28).

(f) (S)- and (R)-(3R,5R) Methyl [3,5-bis(tert.-butyldimethylsilyloxy)-4-hydroxy]-cyclohexylidene carboxylate (5a and 5b).

25 n-Butyllithium (2.3 mL, 3.0 mmol) 1.3 M in hexanes was added to a solution of diisopropylamine (0.42 mL, 3.0 mmol) in anhydrous tetrahydrofuran (2.0 mL) under argon at -78°C with stirring and methyl (trimethylsilyl) acetate (0.49 mL, 3.0 mmol) was added. After 15 min the protected keto compound **4** (0.629 g, 1.4 mmol) in anhydrous tetrahydrofuran (2.0 + 1 mL) was added. The solution was stirred for 2 h at -78°C. The reaction mixture was quenched with saturated ammonium chloride solution and extracted with ether. The combined ether fractions were washed with brine, water and dried over anhydrous MgSO₄, filtered and evaporated. The product was further purified by SepPak filtration in 5% ethyl acetate in hexane to give 0.693 g (98%) of a mixture of the two stereoisomer allylesters **5a** and **5b**. For analytical purposes the two allylesters were separated by HPLC (1% ethyl acetate in hexane, Zorbax Sil 10 x 25 cm, with differential refractometer as detector.)
 35 **5a** Peak II (Major) (S): ¹H NMR (CDCl₃, 500 MHz) δ 0.04, 0.05, 0.08 (3H, 3H and 6H, each s, 4x SiMe), 0.13 (9H, s, SiMe₃), 0.86 (9H, s, Si-t-Bu), 0.89 (9H, s, Si-t-Bu), 2.00 (1H, dd, J=13.5, 4.7 Hz, 6β-H), 2.60 (1H, br d, J=13.5 Hz, 6α-H), 2.74 and 3.28 (1H and 1H, each br m, 2-H₂), 3.62 (1H, narrow m, 4β-H), 3.68 (3H, s, OMe), 3.86 (1H, ~q, J~4 Hz, 5α-H), 3.95 (1H, dt, J=9.5, 2.5 Hz, 3β-H), 5.63 (1H, s, 7-H).
 40 **5b** Peak I (minor) (R): ¹H NMR (CDCl₃, 500 MHz) δ 0.04, 0.05, 0.06 (3H, 3H and 6H, each s, 4x SiMe), 0.13 (9H, s, SiMe₃), 0.84 (9H, s, Si-t-Bu), 0.89 (9H, s, Si-t-Bu), 2.12 (1H, dd, J=12.7, 3.8 Hz, 6α-H), 2.57 (1H, br t, J~12 Hz, 6β-H), 2.62 and 3.35 (1H and 1H, each br d, J~13 Hz, 2-H₂), 3.65 (1H, narrow m, 4α-H), 3.66 (3H, s, OMe), 3.86 (1H, ~q, J~4 Hz, 3β-H), 3.99 (1H, dt, J=9.9, 3.7 Hz, 5α-H), 5.70 (1H, s, 7-H).

45 (g) (S)- and (R)-(3R,5R) [3,5-Bis(tert.-butyldimethylsilyloxy) (4-trimethylsilyloxy)-cyclohexylidene] ethanol (6a and 6b).

A solution of 410 mg of mixture of esters **5a**, **5b** (0.82 mmol) in 8 mL of anhydrous toluene was treated at -78°C under argon with 7 mL (10.5 mmol) of a 1.5 M solution of diisobutylaluminum hydride in toluene. After the addition stirring was continued for 1 h at -78°C. The reaction mixture was then quenched by the addition of 2N potassium sodium tartrate, the organic phase was separated, and the aqueous phase extracted with ethyl acetate. The combined organic phases were washed with water and brine and dried over anhydrous MgSO₄, filtered and evaporated. The residue was purified by fast filtration through a silica gel column, using 20% ethyl acetate in hexane as eluent, to give 352 mg (91%) alcohols **6a** and **6b** which were separated by HPLC, Zorbax Sil 10 x 25 cm column, 10% ethyl acetate in hexane. First eluted the minor 4R alcohol (42 mg) **6b** followed by the major 4S alcohol **6a** (188 mg) (61% recovery).

55 **6a** Peak II (major) (S): ¹H NMR (CDCl₃, 500 MHz) δ 0.04, 0.05, 0.06, 0.07 (3H, each s, 4x SiMe), 0.13 (9H, s, SiMe₃), 0.87 and 0.89 (9H and 9H, each s, 2x Si-t-Bu), 1.93 (1H, dd, J=13.5, 5.5 Hz, 6β-H), 2.24 (1H, br d,

J~12.6 Hz, 2 β -H), 2.38 (1H, dd, J=12.6 Hz, 9.2 Hz, 2 α -H), 2.50 (1H, dd, J=13.5, 3.5 Hz, 6 α -H) 3.57 (1H, narrow m, 4 β -H), 3.80 (1H, dt, J=3.5 Hz, 5.5 Hz, 5 α -H) 3.89 (1H, ddd, J=9.2, 3.0, 2.6 Hz, 3 β -H, 4.13 (2H, m, 8-H₂), 5.45 (1H, t, J=6.9 Hz, 7-H).

6b Peak I (minor) (R): ¹H NMR (CDCl₃, 500 MHz) δ 0.06, 0.07 (6H and 6H, each s, 2x SiMe₂), 0.13 (9H, s, SiMe₃), 0.87 and 0.89 (9H and 9H, each s, 2x Si-t-Bu), 2.06 (1H, dd, J=12.3, 3.9 Hz, 6 α -H), 2.23 (1H, dd, J=13.6, 4.0 Hz, one of 2-H₂), 2.38 (1H, br d, J=13.6 Hz, one of 2-H₂), 2.45 (1H, br t, J~11 Hz, 6 β -H), 3.62 (1H, dd, 4.0 and 2.0 Hz, 4 α -H), 3.82 (1H, ~q, J~4 Hz, 3 β -H), 3.89 (1H, ddd, J=10, 4 and 2 Hz, 5 α -H), 4.03 and 4.12 (1H and 1H, each br m, 8-H₂), 5.57 (1H, t, J=7.1 Hz, 7-H).

10 (h) (S)- and (R)-(3R,5R)-[3,5-Bis(tert.-butyldimethylsilyloxy-4-trimethylsilyloxy)-cyclohexylidene] ethyl di-phenylphosphine oxide (7a and 7b).

265 mg dry allyl alcohol 6a (0.56 mmol) was dissolved in 5.0 mL anhydrous tetrahydrofuran and 0.35 mL (0.56 mmol) n. Butyl lithium (1.6 M in hexanes) added under argon. 108 mg (0.56 mmol) recrystallized dry tosylchloride was dissolved in 1.0 mL anhydrous tetrahydrofuran and added to the allyl alcohol-BuLi solution under argon at 0°C. The solution was stirred at 0°C for 5 min. and set aside at 0°C.

In another dry flask with air replaced by argon 350 μ l (0.56 mmol) n. Butyl lithium (1.6 M in hexanes) was added to 100 μ l (0.56 mmol) diphenylphosphine in 200 μ l anhydrous tetrahydrofuran at 0°C with stirring under argon. To the orange solution was syphoned under argon pressure at 0°C the tetrahydrofuran solution of the allylic tosylate. The resulting orange solution was stirred an additional 30 min at 0°C and quenched by the addition of water. Solvents were evaporated under reduced pressure and the residue was dissolved in 5 mL of dichloromethane. The dichloromethane layer was stirred with 20 mL of 10% hydrogen peroxide at 0°C for 1 h. The dichloromethane layer was separated and washed with cold aqueous sodium sulfite, water and brine, dried over anhydrous MgSO₄, filtered and evaporated. The residue was dissolved in 20% 2-propanol in hexane and passed through a silica SepPak and purified by HPLC (Zorbax-Sil 9.4 x 25 cm column, 20% 2-propanol in hexane) to give 220 mg (60%) of the crystalline phosphine oxide 7a.

7a UV(EtOH): λ_{max} 258, 285, 272 nm. ¹H NMR (CDCl₃, 500 MHz) δ - 0.02, 0.00, 0.01, 0.03 (3H, each s, 4 x SiMe), 0.09 (9H, s, SiMe₃), 0.83 and 0.87 (9H and 9H, each s, 2x Si-t-Bu), 1.86 (1H, br d, J~13.5 Hz, one of 6-H₂), 1.99 and 2.08 (1H and 1H, each m, 2-H₂), 2.42 (1H, br d, J~13.5 Hz, one of 6-H₂), 3.10 (2H, m, 8-H₂), 3.51 (1H, narrow m, 4 β -H), 3.72 (1H, dt, J=3.7, 5.2 Hz, 5 α -H), 3.81 (1H, ddd, J=8.8, 4.2, 2.4 Hz, 3 β -H), 5.24 (1H, q, J=6.9 Hz, 7-H), 7.46, 7.52 and 7.71 (4H, 2H and 4H, each m, Ar-H). Mass spectrum (exact mass calcd for C₃₆H₅₈O₄Si₃P 658.3459 found 658.3453), m/e (relative intensity) 658 (M⁺, 1), 643 (3), 601 (100), 526 (12), 469 (43).

7b was prepared in the same way as 7a except from the corresponding 6b.

7b ¹H NMR (CDCl₃, 500 MHz) δ 0.02, (12 H, s, 2x SiMe), 0.09 (9H, s, SiMe₃), 0.85 (18 H, s, 2x Si-t-Bu), 1.88 (1H, br d, J~14 Hz, one of 2-H₂), 2.01 (1H, br d J~12 Hz, 6 α -H), 2.06 (1H, br d, J~14 Hz, one of 2-H₂), 2.34 (1H, br m, 6 β -H) 3.02 and 3.14 (1 H and 1 H, each br m, 8-H₂), 3.51 (1H, narrow m, 4 α -H), 3.71 (1H, ~q, J=4.4 Hz, 3 β -H), 3.84 (1H, m, 5 α -H), 5.27 (1H, m, 7-H), 7.46, 7.52 and 7.71 (4H, 2H and 4H, each m, Ar-H).

40 (i) Triethylsilyloxy Grundmann Ketone (8).

Referring now to Scheme II, the CD-ring fragment for the 19-nor vitamin D-derivative was prepared by ozonolysis of commercial vitamin D₃ to provide 20 followed by RuO₄ oxidation to give the 25-hydroxy-Grundmann ketone 21. To 30 mg ketone 21 (0.1 mmol) and 28 mg imidazole (0.41 mmol) in 500 μ l anhydrous dimethyl formamide was added 40 μ l triethylsilyl chloride (0.24 mmol). The mixture was stirred at room temperature for 2 h. Ethyl acetate was added and water, and separated. The ethyl acetate layer was washed with water, brine, dried over anhyd. MgSO₄, filtered and evaporated. The residue was passed through a silica gel SepPak column in 10% ethyl acetate in hexane, and after evaporation purified by HPLC (Zorbax Sil 9.4 x 25 cm column, 10% ethyl acetate in hexane, using a differential refractometer) to give 31 mg (79%) of the pure protected ketone 8.

¹H NMR (CDCl₃, 500 MHz) δ 0.56 (6H, q, J=8.0 Hz, 3 x Si-CH₂), 0.64 (3H, s, 18-H₃), 0.94 (9H, t, J=8.0 Hz, 3 x SiEt), 0.95 (3H, d, J=6.5 Hz, 21-H₃), 1.19 (6H, br s, 26- and 27-H₃), 2.45 (1H, dd, J=11.7, 7.4 Hz, 14 α -H).

65 (j) 1 α ,2 α ,25-trihydroxy-19-nor-vitamin D₃ (10a).

16.9 mg (25.7 μ mol) phosphine oxide 7a was dissolved in 200 μ l anhydrous tetrahydrofuran, cooled to 0°C and 20 μ l (26 μ mol) n.butyl lithium (1.3 molar in hexanes) added under argon with stirring. The solution turns deep orange. The mixture was cooled to -78°C and 7.5 mg (21 μ mol) protected ketone 8 added in 200

5 μl + 100 μl anhydrous tetrahydrofuran. The mixture was stirred under argon at -78°C for 1 h (at that time the solution became colorless) and at RT 18 hr. Ethyl acetate was added and the organic phase washed with water, brine, dried over anhydrous MgSO_4 , filtered and evaporated. The residue was dissolved in 10% ethyl acetate in hexane, passed through a silica SepPak and washed with 40 mL of the same to give the 19-nor vitamin D derivative **9a**. The Sep Pak was then washed with 20% 2-propanol in hexane to recover 5 mg unchanged di-phenylphosphine oxide. **9a** was purified by HPLC in 10% ethyl acetate in hexane (Zorbax Sil 9.4 x 25 cm column) to give 8.2 mg of the protected 19-nor vitamin D₃ derivative **9a** (54%).

10 ¹H NMR (CDCl_3 , 500 MHz) δ 0.04, 0.05, 0.06 (3H, 3H, and 6H, each s, 4x SiMe₃), 0.12 (9H, s, SiMe₃), 0.55 (3H, s, 18-H₃), 0.56 (6H, q J=7.4 Hz, 3 x SiCH₂), 0.87 and 0.88 (9H and 9H, each s, 2 x Si-t-Bu), 0.92 (3H, d, J=6.1 Hz, 21-H₃), 0.95 (9H, t, J=7.4 Hz, 3 x SiEt), 1.19 (6H, br s, 26- and 27-CH₃), 2.79 (1H, br d, J=12.6 Hz, 9 β -H), 3.53 (1H, m, 2 β -H), 3.80 (1 H, m, 3 α -H), 3.88 (1 H, m, 1 β -H), 5.81 and 6.10 (1 H and 1 H, each d, J=11.4 Hz, 6- and 7-H). Mass spectrum (exact mass calcd for C₄₇H₉₄O₄Si₄ 834.6229, found 834.6241) m/e (relative intensity) 834 (12), 805 (3), 702 (100), 645 (18), 599 (45).

15 All of **9a** was dissolved in 1.0 ml of anhydrous tetrahydrofuran and treated with 150 μl of a 1M solution of tetrabutylammonium fluoride in tetrahydrofuran. The mixture was stirred under argon at RT for 16 h and extracted with ethyl acetate. The organic phase was washed with 10% NaHCO₃ solution, brine and dried over anhydrous MgSO_4 , filtered and evaporated. The residue was filtered through a Silica SepPak and purified by HPLC (Zorbax Sil 9.4 x 25 cm column, 30% 2-propanol in hexane) to give 1.48 mg of pure 1 α ,2 α ,25-trihydroxy-19-nor-vitamin D₃ **10a**.

20 UV (In EtOH) λ_{max} : 243, 251.5, 261 nm. Mass spectrum m/e (relative intensity): 420 (M⁺, 100), 402 (55), 384 (18), 245 (45), 95 (95), 59 (95).

10a (2 α -OH): ¹H NMR (CDCl_3 , 500 MHz) δ 0.55 (3H, s, 18-H₃), 0.94 (3H, d, J=6.5 Hz, 21-H₃), 1.22 (6H, s, 26- and 27-CH₃), 2.62 (1H, dd, J=13.0, 4.2 Hz), 2.79 (1H, br d, J=12.9 Hz), 2.90 (1H, dd, J=14.3, 4.2 Hz), 3.25 (1H, br m), 3.53 (1H, dd, J=8.4, 2.6 Hz), 3.79 (1 H, br m), 4.10 (1 H, m), 5.81 and 6.38 (1 H and 1 H, each d, J=11.0 Hz, 6- and 7-H).

25 **10b** was prepared in the same way as **10a** except from the corresponding **7b** (Structure not shown).

10b (2 β -OH): ¹H NMR (CDCl_3 , 500 MHz) δ 0.55 (3H, s, 18-H₃), 0.94 (3H, d, J=6.8 Hz, 21-H₃), 1.22 (6H, s, 26- and 27-CH₃), 2.79 (1H, br d, J=13.0 Hz), 3.36 (2H, br m), 3.49 (1H, m), 3.68 (1 H, br m), 4.08 (1 H, m), 5.84 and 6.29 (1 H and 1 H, each d, J=11.3 Hz, 6- and 7-H).

30 Example 2

Scheme III illustrates the preparation of 1 α ,25-dihydroxy-2 α (3'-hydroxypropoxy)-19-nor-vitamin D₃ (**14a**) and 1 α ,25-dihydroxy-2 α (benzyloxy)-19-nor-vitamin D₃ (**16a**).

35 (a) 19-Nor-(1 α ,3 β)-[bis(tert.-butyldimethylsilyloxy)-25-triethylsilyloxy]-2 α -hydroxyvitamin D₃ (**11a**).

40 10 mg **9a** was stirred for 4.5 h at room temperature with 3 ml of a mixture of tetrahydrofuran-acetic acid-water (8:8:1). Ethyl acetate was added and washed with ice cold water, ice cold 10% NaHCO₃ solution until neutral, water and brine, and dried over anh. MgSO_4 , filtered and evaporated. The residue was purified by HPLC (Zorbax Sil 10 x 25 cm column, 5% ethyl acetate in hexane) to give (in order of peak elutions) 1.2 mg unchanged starting material (12%), 3.5 mg of the expected 2-hydroxy compound **11a** (38%) and 0.75 mg of 25-hydroxy compound (10%) (total recovery 60%).

45 **11a** (2 α -OH): ¹H NMR (CDCl_3 , 500 MHz) δ 0.06, 0.07, 0.08, 0.10 (each 3H, each s, 4x SiMe₃), 0.55 (3H, s, 18-H₃), 0.56 (6H, q, J=7.8 Hz, 3x SiCH₂), 0.87 and 0.88 (9H and 9H, each s, 2 x Si-t-Bu), 0.93 (3H, d, J=6.4 Hz, 21-H₃), 0.95 (9H, t, J=7.8 Hz, 3 x SiEt), 1.19 (6H, br s, 26- and 27-CH₃), 2.27 (1H, d, J=3.3 Hz, OH), 2.79 (1H, dd, J=12.3, 4.2 Hz, 9 β -H), 3.51 (1H, dt, J=6 and 3 Hz; after D₂O : dd, J=5.8, 2.7 Hz, 2 β -H), 3.91 (1H, dt, J=4.3, 5.8 Hz, 3 α -H), 4.00 (1H, ~dt, J=7.5, 3 Hz, 1 β -H), 5.80 and 6.16 (1H and 1H, each, d, J=11.2 Hz, 6- and 7-H).

19-Nor-(1 α ,3 β)-[bis (tert.-butyldimethylsilyloxy)-25-triethylsilyloxy]-2 β -hydroxy-vitamin D₃ (11b**)**

50 **11b** (not shown) was prepared the same was as **11a** except from **9b** (not shown)

11b (2 β -OH): ¹H NMR (CDCl_3 , 500 MHz) δ 0.06, 0.07, 0.08, 0.10 (each 3H, each s, 4x SiMe₃), 0.54 (3H, s, 18-H₃), 0.56 (6H, q, J=8.0 Hz, 3x SiCH₂), 0.86 and 0.89 (9H and 9H, each s, 2x Si-t-Bu), 0.93 (3H, d, J=6 Hz, 21-H₃), 0.94 (9H, t, J=8.0 Hz, SiEt), 1.19 (6H, br s, 26- and 27-CH₃), 2.81 (1H, br d, J=13 Hz, 9 β -H), 3.59 (1H, narrow m, after D₂O : dd, J=3.6, 3.3 Hz, 2 α -H), 4.00 (2H, m, 1 β - and 3 α -H), 5.80 and 6.19 (1H and 1H, each d, J=11.0 Hz, 6- and 7-H).

(b) 3-Bromo-1-(tert.-butyldimethylsilyloxy)-propane (12).

1.4 g (1 mmol) 3-bromo-1-propanol was dissolved in 5 ml of anhydrous dimethyl formamide and 3.0 g of imidazole, followed by 3.3 g of tert.-butyldimethylsilyl chloride was added at 0°C with stirring. The mixture was stirred at room temperature for 2 h. Ether was added, and the ether phase washed with water and brine, dried over anhydrous MgSO_4 , filtered and evaporated. The residue was passed through a small silica gel column and eluted with hexane to give 2.03 g (80%) pure **12**.

^1H NMR (CDCl_3 , 500 MHz) δ 0.06 (6 H, s, SiMe_2), 0.90 (9H, s, Si-t-Bu), 2.03 (2H, ~ quint., $J=6$ Hz, C- CH_2 -C), 3.51 (2H, t, $J=6.2$ Hz, CH_2 -O), 3.73 (2H, t, $J=5.8$ Hz, CH_2 -Br).

(c) 19-Nor-2 α -(3'-hydroxypropoxy)-1-dihydroxyvitamin D₃ (14a).

1.6 mg (2 μmol) **11a** was dissolved in 200 μl of anhydrous dimethyl formamide and 3 mg sodium hydride (as 60% oil dispersion) followed by 3 mg 18-Crown-6 and 5 μl of the bromo compound **12** was added and the mixture stirred under argon atmosphere for 48 h. The mixture was extracted with ethyl acetate, washed with water, dried over anhydrous MgSO_4 , filtered and evaporated. The residue was passed through a silica gel SepPak column and evaporated in AcOEt.

13a: ^1H NMR (CDCl_3 , 500 MHz) δ 0.04, 0.05, 0.06, 0.07 (6H, 6H, 3H, 3H, each s, 6x SiMe), 0.55 (3H, s, 18- H_3), 0.56 (6H, q, $J=7.5$ Hz, 3x SiCH_2) 0.87 and 0.88 and 0.89 (9H, 9H and 9H, each s, 3x Si-t-Bu), 0.93 (3H, d, $J=6$ Hz, 21- H_3), 0.95 (9H, t, $J=7.5$ Hz, 3 x SiEt), 1.19 (6H, br s, 26-and 27- CH_3), 2.79 (1H, br d, $J=14$ Hz), 3.12 (1H, m), 3.4-4.1 (at least 7H, complex m), 5.80 and 6.12 (1H and 1H, each d, $J=11$ Hz, 6- and 7-H).

The residue was dissolved in 1 ml of anhydrous tetrahydrofuran and 0.5 ml tetrabutylammonium fluoride (1.0 M in THF) was added and stirred under argon atmosphere for 20 h. The residue was extracted with ethyl acetate, washed with water and 10% NaHCO_3 solution, water and brine, and dried over anhydrous MgSO_4 , filtered and evaporated. The residue was passed through a silica gel SepPak column in 1:1 2-PrOH-hexane and purified by HPLC (Zorbax Sil 10mm x 25 cm column, 40% 2-PrOH-hexane) to give 202 μg of the expected product **14a** (overall yield from **11a** 21%) and 20 μg of **10a**.

14a UV (in EtOH) λ_{max} : 243, 251.5, 261 nm. Mass spectrum (exact mass calcd for $\text{C}_{29}\text{H}_{50}\text{O}_8$ 478.3658, found 478.3659), m/e (relative intensity) 478 (M^+ , 5), 460 (6), 442 (2), 402 (4), 384 (3), 245 (15), 184 (20), 142 (100), 95 (50), 59 (38). ^1H NMR (CDCl_3 , 500 MHz) δ 0.55 (3H, s, 18- H_3), 0.93 (3H, d, $J=6.8$ Hz, 21- H_3), 1.22 (6H, s, 26-and 27- H_3), 3.3-4.2 (at least 7H, complex m), 5.83 and 6.34 (1H and 1H, each d, $J=11.2$ Hz, 6- and 7-H).

(d) 19-Nor-2 β -(3'-hydroxypropoxy)-1 α ,25-dihydroxyvitamin D₃ (14b) (Structure not shown)

14b was prepared in the same way as **14a** except from the corresponding **11b** (structure not shown).

14b UV (in EtOH) λ_{max} : 243, 251.5, 261 nm. Mass spectrum m/e (relative intensity) 478 (M^+ , 5), 460 (20), 442 (18), 424 (4), 384 (3), 245 (28), 181 (28), 95 (50), 69 (100), 59 (68). ^1H NMR (CDCl_3 , 500 MHz) δ 0.54 (3H, s, 18- H_3), 0.94 (3H, d, $J=6.5$ Hz, 21- H_3), 1.22 (6H, s, 26-and 27- H_3), 2.79 (1H, br d, $J=12$ Hz, 9 β -H), 3.0-4.3 (at least 7H, complex m), 5.83 and 6.29 (1H and 1H, each d, $J=11.2$ Hz, 6- and 7-H).

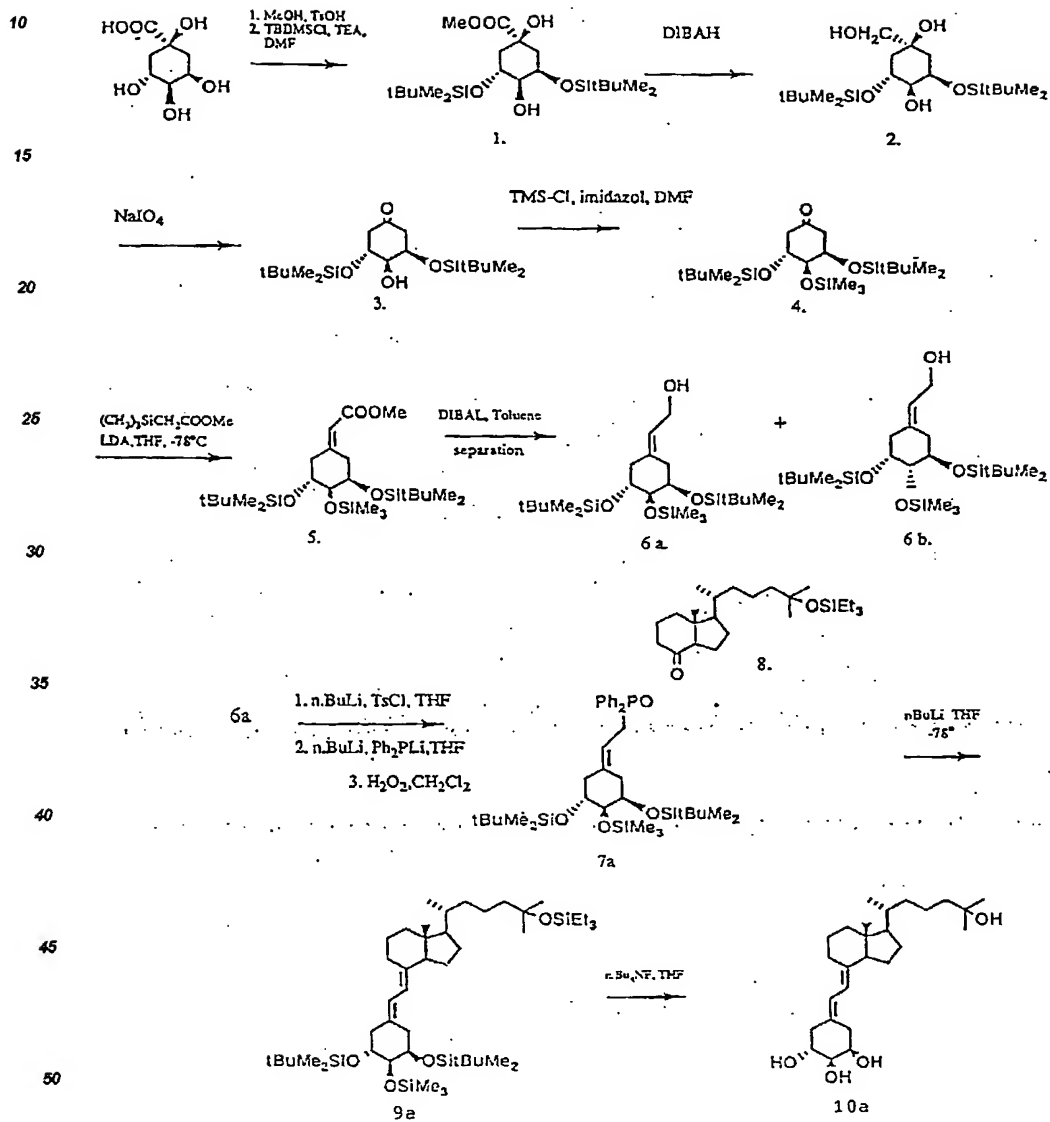
(e) 19-Nor-2 α -(benzyloxy)-1 α ,25-dihydroxyvitamin D₃ (16a).

1.6 mg (2 μmol) (**11a**) was dissolved in 200 μl of anhydrous dimethyl formamide and 3 mg sodium hydride (as 60% oil dispersion) followed by 3 mg 18-Crown-6 and 6 μl of benzylbromide benzene solution, (prepared from 120 μl benzylbromide in 1 mL benzene) was added and the mixture stirred under argon atmosphere for 48 h. The mixture was extracted with ethyl acetate, washed with water, dried over anhydrous MgSO_4 , filtered and evaporated. The residue was passed through a silica gel SepPak column, evaporated to give 860 μg crude **15a**, which was without purification deprotected. 860 μg crude **15a** was dissolved in 200 μL of methanol and 10 mg methanol washed AG-50W-X4 cation exchange resin added. The mixture was stirred under argon atmosphere at RT for 18 h, filtered through a SepPak Silica cartridge and washed with 2-propanol. The solvent was evaporated under reduced pressure and the residue purified by HPLC (Zorbax Sil 10 mm x 25 cm column, 30% 2-PrOH-hexane mixture) to give 170 μg of the 2 α -benzyloxy-compound mixture to give 170 μg of the 2 α -benzyloxy-compound **16a**.

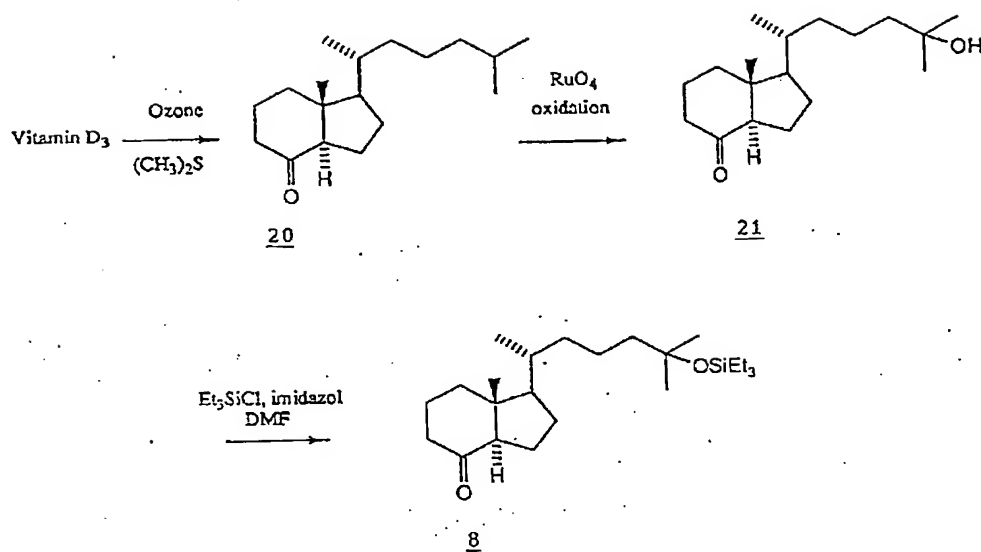
16a: UV (in EtOH) λ_{max} : 243, 251.5, 261 nm. Mass spectrum (exact mass calcd for $\text{C}_{33}\text{H}_{50}\text{O}_4$ 510.3709 found 510.3703), m/e relative intensity 510 (11), 492 (8), 474 (2), 401 (8), 91 (100). ^1H NMR (CDCl_3 , 500 MHz) δ 0.55 (3 H, s, 18- H_3), 0.93 (3 H, d, $J=6.7$ Hz, 21- H_3), 1.22 (6H, s, 26- and 27- H_3), 2.79 (2H, m), 3.45 (1 H, dd, $J=7.3$, 3.0 Hz, 2 β -H), 3.97 (1 H, m, 3 α -H), 4.11 (1 H, m, 1 β -H), 4.65 and 4.72 (1 H and 1 H, each d, $J=11.8$ Hz, O- CH_2), 5.83 and 6.33 (1 H and 1 H, each d, $J=11.2$ Hz, 6- and 7-H), 7.2-7.4 (5H, br m, Ar-H).

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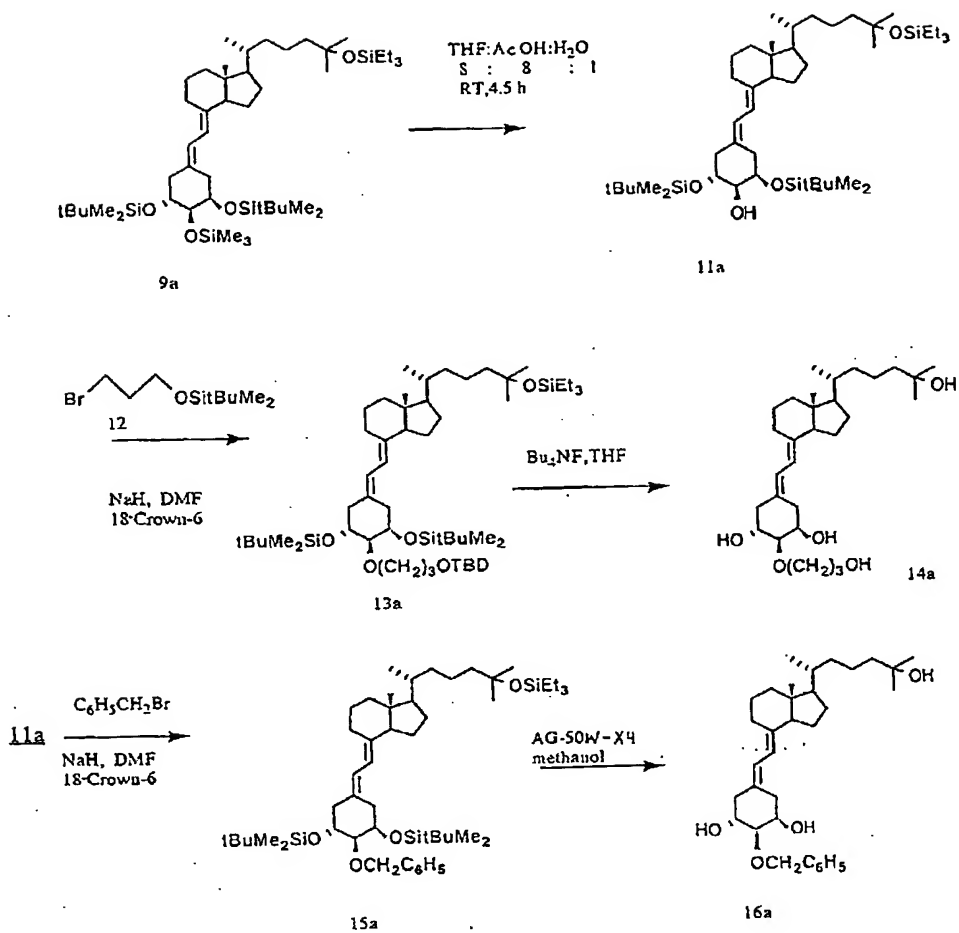
SCHEME I



SCHEME II



SCHEME III



SCHEME IV

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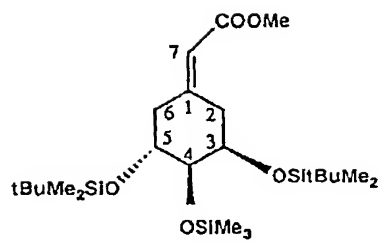
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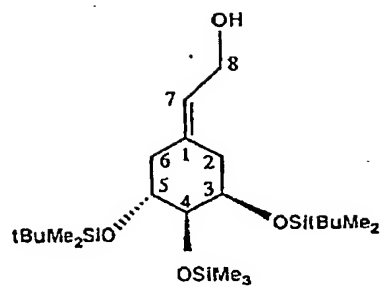
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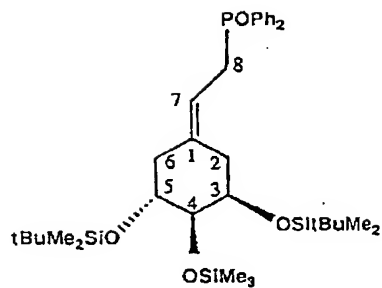
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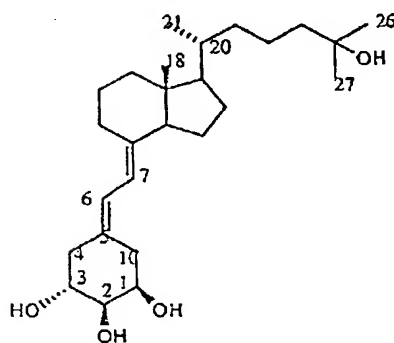
5a.



6 a.



7a



10a

Biological Activity of 1 α -Hydroxy-19-Nor Vitamin D Compounds

The novel compounds of this invention exhibit an unexpected pattern of biological activity. All of the 19-nor compounds exhibited high potency in promoting the differentiation of malignant cells. The two 2-hydroxy analogs showed in vivo calcium transport with little or no bone calcium mobilization; the 2 β - more than the 2 α - analog; while the 2 α -hydroxypropoxy analog showed a selective activity profile, combining high potency in inducing differentiation of malignant cells with very low bone calcium mobilizing activity. This is illustrated by the biological assay results obtained for the claimed 19-nor-vitamin D₃ compounds, which are summarized in Figs. 1 and 2, and Table 1. Fig. 1 shows a comparison of the activity of the known active metabolite 1 α ,25-dihydroxyvitamin D₃ and two of the 19-nor analogs in inducing the differentiation of human leukemia cells (HL-60 cells) in culture to normal cells (monocytes). Differentiation activity was assessed by a standard differentiation assay, abbreviated in Fig. 1 as NBT (nitroblue tetrazolium reduction). The assay was conducted according to known procedures, as given, for example, by DeLuca et al U. S. Patent 4,717,721 and Ostrem et al, J. Biol. Chem. 262, 14164, 1987. For the assay, the differentiation activity of the test compounds is expressed in terms of the percent of HL-60 cells having differentiated to normal cells in response to a given concentration of test compound.

The results summarized in Fig. 1 clearly show that the new analogs, 1 α ,2 α ,25-trihydroxy-19-nor-vitamin D₃ and 1 α ,25-dihydroxy-2 α -(3'-hydroxypropoxy)-19-nor-vitamin D₃, are as potent as 1 α ,25-dihydroxyvitamin D₃ in promoting the differentiation of leukemia cells. Thus in NBT assay close to 90% of the cells are induced to differentiation by 1 α ,25-dihydroxyvitamin D₃ at a concentration of 1 x 10⁻⁷ molar, and the same degree of differentiation is achieved by the two 19-nor analogs.

Fig. 2 shows a comparison of the same three compounds as in Fig. 1 illustrating their relative activity with regard to competitive binding to the vitamin D receptor. The competitive receptor binding was done with pig nuclear extract as described in Perlman et al. Biochemistry 29, 190-196 (1990) using the porcine extract prepared as described by Dame et al PNAS 82, 7825-7829 (1985). These data are used to demonstrate that the compounds described herein have relatively high *in vivo* activity, and have somewhat less activity than 1,25-(OH)₂D₃ in binding to the vitamin D receptor.

In regard to the biological data on the calcemic activity of these compounds reported in Table 1, Holtzmann weanling rats were maintained on 0.47% Ca, 3% P, for one week, then switched to a low Ca diet (0.02% Ca) for an additional three weeks. During the 4th week all animals were dosed with the appropriate compounds via the peritoneal cavity. All doses were suspended in ethanol propylene glycol (5/95) and administered daily for seven days. None of the compounds produced hypercalcemia over the seven day dosing period. The 19-nor-2 α -hydroxypropoxy-1 α ,25-dihydroxyvitamin D₃ did mobilize small amounts of Ca from bone at 130 or 325 pmole daily. However, these levels could easily be controlled by lowering the dose and the activity in this regard was far below that of the standard compound, 1,25-(OH)₂D₃.

The data in Table 1 illustrates that 19-nor-1 α ,2 α ,25-trihydroxyvitamin D₃ has biological activity in intestinal transport similar to that of 1,25-(OH)₂D₃ but possesses little or no bone calcium mobilizing activity even when given at 325 pmol/day. Similarly, the 1 α ,2 β ,25-trihydroxyvitamin D₃ compound has considerable activity in intestinal calcium transport but again lacks bone calcium mobilizing activity. On the other hand, the 19-nor-2 α -hydroxypropoxy-1 α ,25-dihydroxyvitamin D₃ compound has an activity profile similar to 1,25-(OH)₂D₃ but has preferential activity on bone calcium mobilization.

The 2-propoxy compound would be useful in circumstances where an increased bone turnover is desirable such as low bone turnover osteoporosis. The other two compounds would find utility as a treatment for post-menopausal and senile osteoporosis because of their low bone calcium mobilizing activity with normal differentiative activity, normal binding to the receptor, and normal calcium transport activity. The compounds that could be considered for anti-cancer or psoriasis might be the 2 α - and 2 β -hydroxy compounds because of their tendency to not cause hypercalcemia.

For treatment purposes, the novel compounds of this invention can be formulated as solutions in innocuous solvents, or as emulsions, suspensions or dispersions in suitable innocuous solvents or carriers, or as pills, tablets or capsules, containing solid carriers according to conventional methods known in the art. For topical applications the compounds are advantageously formulated as creams or ointments or similar vehicle suitable for topical applications. Any such formulations may also contain other pharmaceutically-acceptable and non-toxic excipients such as stabilizers, anti-oxidants, binders, coloring agents or emulsifying or taste-modifying agents.

The compounds are advantageously administered by injection, or by intravenous infusion of suitable sterile solutions, or in the form of oral doses via the alimentary canal, or topically in the form of ointments, lotions, or in suitable transdermal patches. For the treatment of malignant diseases, the 19-nor vitamin D compounds of this invention are administered to subjects in dosages sufficient to inhibit the proliferation of malignant cells

and induce their differentiation into normal monocyte-macrophages. Suitable dosage amounts are from 1 to 500 µg of compound per day, as a single dose or individual dosages, such dosages being adjusted, depending on the disease to be treated, its severity and the response or condition of the subject as is well-understood in the art.

It can thus be seen that the compounds of this invention find utility in the treatment of senile or post menopausal osteoporosis as well as in low bone turnover osteoporosis. The compounds can be administered either during and subsequent to menopause or prior to the onset of menopause.

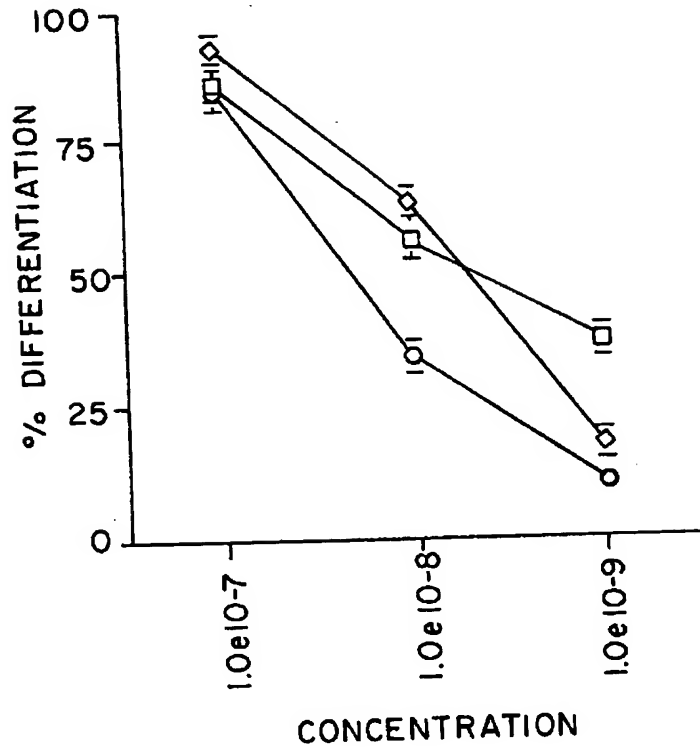
TABLE 1

| Group | Amount pmoles/d/ 7 days | S/M Ratio ave. s.e.m. | Serum Ca ave. s.e.m. (mg %) |
|---|-------------------------------|--|--|
| D-Deficient | 0 | a2.1 ± 0.18 | a4.3 ± 0.14 |
| 1α, 25-(OH) ₂ D ₃ | 130 | b8.0 ± 0.80 | b5.6 ± 0.20 |
| 19-Nor-1α, 2α, 25-(OH) ₃ D ₃ | 130 | c5.3 ± 0.26 | 1ca4.12 ± 0.3 |
| | 325 | 5.0 ± 0.14 | cb4.14 ± 0.10 |
| D-Deficient | 0 | a3.0 ± 0.8 | a4.3 ± 0.23 |
| 19-Nor-1α, 2β, 25-(OH) ₃ D ₃ | 65 | c5.8 ± 0.22 | 2ca4.2 ± .19 |
| | 325 | 10.8 ± 0.39 | cb4.2 ± 0.28 |
| | 650 | 9.9 ± 0.45 | ca4.3 ± 0.24 |
| 1α, 25-(OH) ₂ D ₃ | 65 | b8.3 ± 0.32 | b15.2 ± 0.16 |
| | 325 | 10.5 ± 1.2 | b26.8 ± 0.24 |
| D-Deficient | 0 | a2.5 ± 0.15 | a4.3 ± 0.13 |
| 1α, 25-(OH) ₂ D ₃ | 130 | b7.1 ± 0.49 | b15.7 ± 0.23 |
| | 325 | 7.8 ± 0.60 | b27.1 ± 0.30 |
| 19-Nor- 2α-hydroxy- propoxy- 1α, 25-(OH) ₂ D ₃ | 130 | c3.5 ± 0.23 | 3ca4.9 ± 0.11 |
| | 325 | 4.3 ± 0.33 | 3b5.8 ± 0.20 |
| Statistical data | | | |
| Serosal/Mucosal | | Serum Ca | |
| Panel 1 All from a | P < .001 | Panel 1, b from a P < .001 | 1ca, cb from a N.S. |
| Panel 2 All from a | P < .001 | Panel 2, b ¹ , b ² , from a p<.001 | 2ca ¹ cb ² cc N.S. |
| Panel 3 All from a | P < .001 | Panel 3 b ¹ , b ² from a p < .001 | (3) ca from a p < .0005 |
| | | | cb from a, p< .001 |

Claims

1. A vitamin D compound having the structure:

HL-60 DIFFERENTIATION BY NBT ASSAY



- 1α,25(OH)₂D₃
- ◇— 19 NOR-1α25(OH)₂-2α(3 OH-PROPOXY)D₃
- 19 NOR-1α2α25(OH)₃D₃

FIG. 1A

HL-60 CELL DIFFERENTIATION - NBT ASSAY

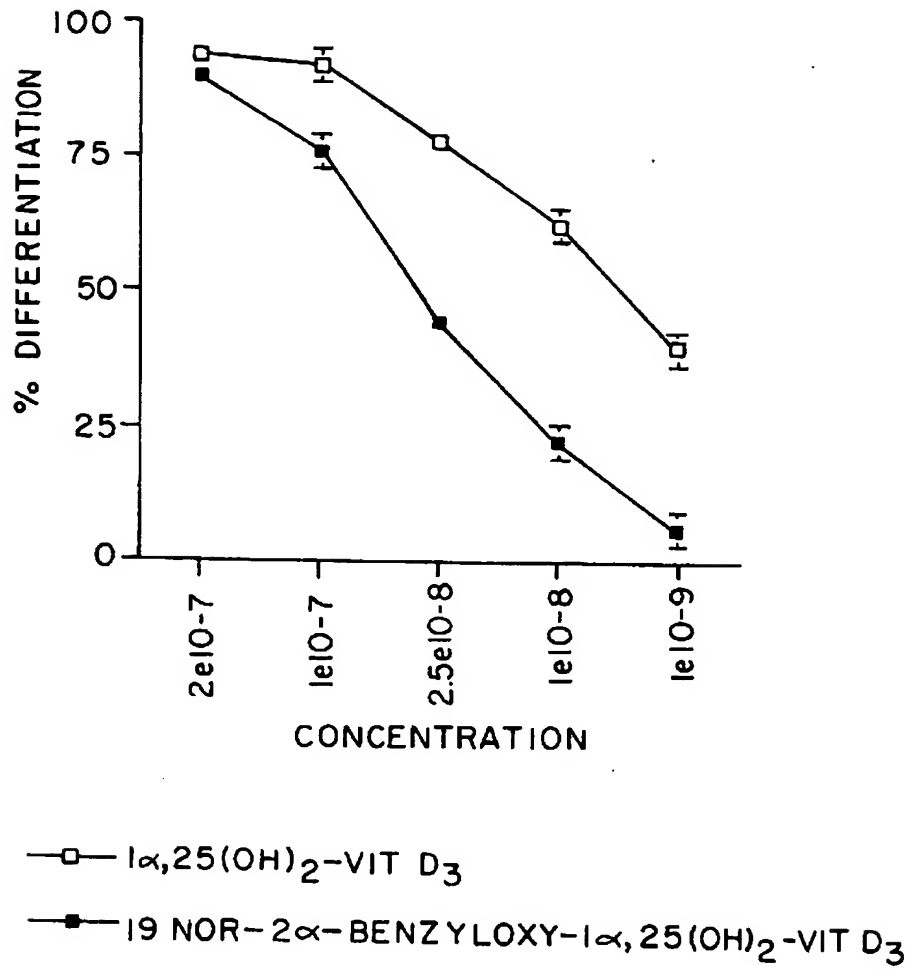
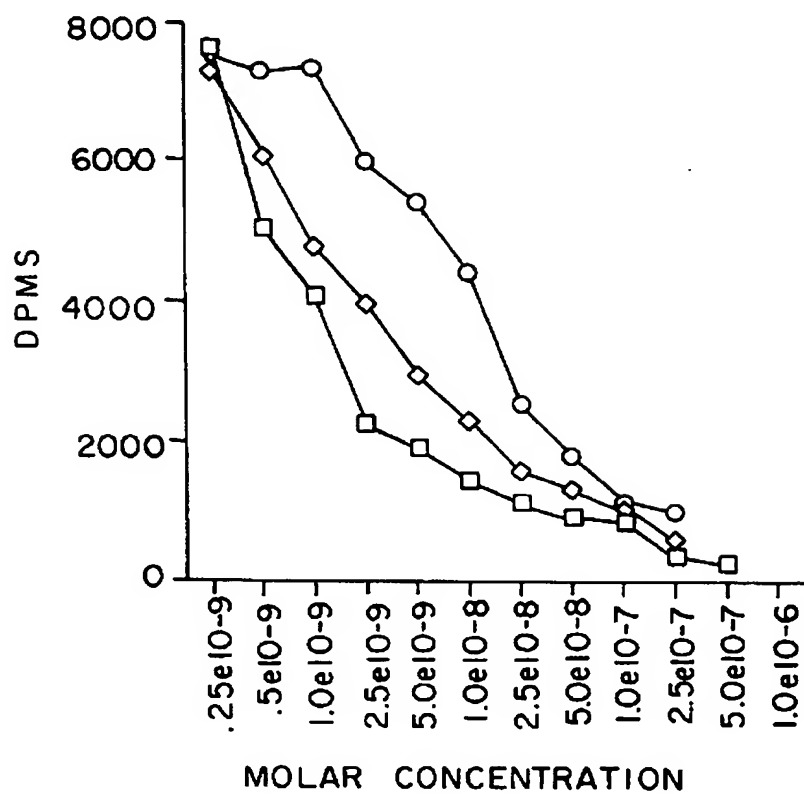


FIG. 1B

COMPETITIVE BINDING TO VITAMIN D RECEPTOR



- $1\alpha,25(\text{OH})_2 \text{ VIT D}_3$
 —◇— $19\text{NOR}-1\alpha,25(\text{OH})_2-2\alpha(3\text{OH-PROPOXY}) \text{ VIT D}_3$
 —○— $19\text{NOR}-1\alpha,2\alpha,25(\text{OH})_3 \text{ VIT D}_3$

FIG. 2A

COMPETITIVE BINDING TO VDR-PNE

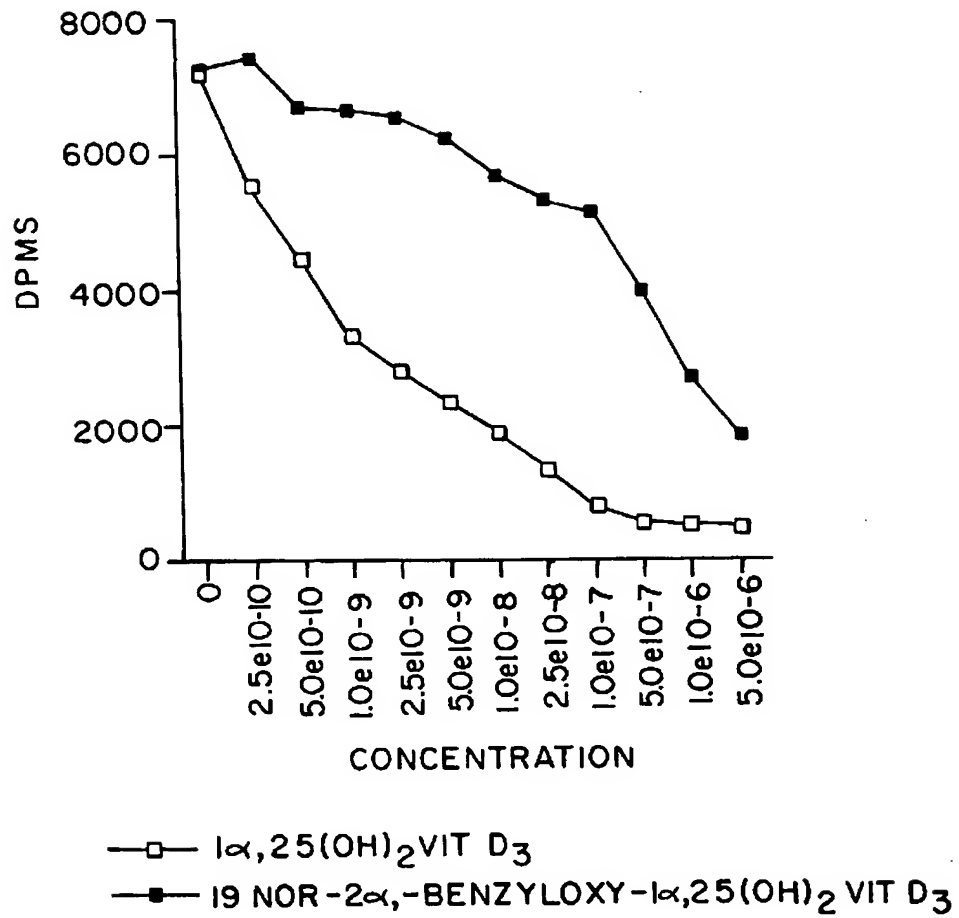


FIG. 2B



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Application Number
EP 94 30 2382

| DOCUMENTS CONSIDERED TO BE RELEVANT | | | |
|--|---|---|--|
| Category | Citation of document with indication, where appropriate, of relevant passages | Relevant to claim | CLASSIFICATION OF THE APPLICATION (Int. CL.5) |
| A | EP-A-0 387 077 (WISCONSIN ALUMNI RESEARCH FOUNDATION) * page 7, line 55 - page 8, line 24; examples 1e, 2 e; claims 1-4, 12-18 * | 1,7 | C07C401/00 A61K31/59 |
| A | EP-A-0 516 410 (WISCONSIN ALUMNI RESEARCH FOUNDATION) * page 15, compound 20 * | 1 | |
| A | EP-A-0 184 206 (CHUGAI SEIYAKU KK) * page 1, lines 3-10; examples 1-4, 6, 7, 9, 10, 13, 14; claims 1-5, 7 * & US-A-4 666 634 | 1,5,7,8, 12 | |
| D | | | |
| A | J. ORG. CHEM., vol.56, no.14, 1991 pages 4339 - 4341 G.H. POSNER ET AL * page 4339, summary * | 1,2 | |
| A | CHEMICAL ABSTRACTS, vol. 110, no. 10, 1989, Columbus, Ohio, US; abstract no. 82505y, * abstract * & JP-A-63 107 929 (CHUGAI PHARMACEUTICAL CO LTD) | 1,5,7-9, 12 | TECHNICAL FIELDS SEARCHED (Int. CL.5) C07C A61K |
| The present search report has been drawn up for all claims | | | |
| Place of search BERLIN | | Date of completion of the search 24 May 1994 | Examiner Van Amsterdam, L |
| <p>CATEGORY OF CITED DOCUMENTS</p> <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p> <p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons A : member of the same patent family, corresponding document</p> | | | |

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